Amendments to the Specification:

Amend the paragraph beginning at page 18, line 12, of the English language specification, as follows.

PCR amplification was conducted using pSA212 as a template and using primers 5-1F (SEQ ID NO: 9) and 5-1R (SEQ ID NO: 10) for SIVagmTYO1-derived 5'LTR region (8547-9053+1-982, which was added KpnI site at the 5' end and EcoRI site at the 3' end); primers 5-2F (SEQ ID NO: 11) and 5-2R (SEQ ID NO: 12) for RRE (~~7380~~7370-7993, which was added EcoRI site at the 5' end and SacII site at the 3' end); or primers 5-3F (SEQ ID NO: 13) and 5-3R (SEQ ID NO: 14) for 3'LTR (8521-9170, which was added NotI and BamHI sites at the 5' end, and SacI site at the 3' end). CMV promoter and EGFP encoding region (1-1330; which was added SacII site at the 5' end, and added NotI site and BamHI site as well as a translational stop codon at the 3' end) derived from pEGFPC2 (Clontech) was amplified by PCR using primers 6F (SEQ ID NO: 15) and 6R (SEQ ID NO: 16), and pEGFPC2 as a template. The four types of PCR fragments were respectively digested with a pair of restriction enzymes of KpnI and EcoRI, a pair of EcoRI and SacII, a pair of BamHI and SacI, and a pair of SacII and BamHI, followed by purification. Then, they were ligated in the order of 5'LTR, 3'LTR, RRE and CMV promoter EGFP prior to the insertion between KpnI-SacI site of pBluescript KS+ (pBS/5'LTR.U3G2/RREc/s/CMVFEGFP/WT3'LTR). When β-galactosidase was used as a reporter gene, the DNA fragments containing the 5'LTR region and 3' LTR region respectively were prepared by PCR as described above. After digestion with a pair of restriction enzymes KpnI and EcoRI and a pair of NotI and SacI respectively, the DNA

fragments were purified, and then inserted at the KpnI-EcoRI site and the NotI-SacI site of pBluescript KS+, respectively (pBS/5' LTR.U3G2/WT3' LTR). A NotI fragment containing the region encoding β-galactosidase of pCMV-beta (Clontech) (820-4294) was inserted into the plasmid at the NotI site (pBS/5' LTR.U3G2/beta-gal/WT3' LTR). Then, an RRE sequence (6964-8177; which was added EcoRI site at the 5' end and added NotI site at the 3' end), which had been amplified by PCR using primers 7-1F (SEQ ID NO: 17) and 7-1R (SEQ ID NO: 18) as well as using pSA212 as a template, was inserted at the EcoRI-NotI site in plasmid pBS/5' LTR.U3G2/beta-gal/WT3' LTR (pBS/5' LTR.U3G2/RRE6/tr/beta-gal/WT3' LTR). The RRE sequence was cut out with EcoRI and NheI prior to the insertion of the RRE sequence (~~7380~~7370-7993; which was added EcoRI site at the 5' end and added NheI site at the 3' end), which had been amplified by PCR using primers 7-2F (SEQ ID NO: 19) and 7-2R (SEQ ID NO: 20) as well as using pSA212 as a template. After the resulting plasmid (pBS/5' LTR.U3G2/RREc/s/beta-gal/WT3' LTR) was digested with NheI and SmaI and blunted, a CMV promoter region (8-592; blunted AseI-NheI fragment) derived from pEGFPN2 (Clontech) was inserted therein (pBS/5' LTR.U3G2/RREc/s/CMVFbeta-gal/WT3' LTR). All blunting reactions were performed using a Blunting High (Toyobo) according to the attached instruction. The plasmids pBS/5' LTR.U3G2/RREc/s/CMVFEGFP/WT3' LTR and pBS/5' LTR.U3G2/RREc/s/CMVFbeta-gal/WT3' LTR were digested with KpnI and SacI respectively to provide DNA fragments containing the region between 5' LTR-3' LTR. The fragments were inserted into pGL3 Control vector (Promega) at the KpnI-SacI site for use as a gene transfer vector (pGL3C/5' LTR.U3G2/RREc/s/CMVFbeta-gal or

EGFP/WT3' LTR). For the identification of packaging signal, the 5'LTR region was cut off with KpnI and EcoRI from pBS/5' LTR.U3G2/RREc/s/CMVFbeta-gal/WT3' LTR plasmid, and a variety of DNA fragments were prepared for each containing a region of different length by PCR using a primer 8F (SEQ ID NO: 21) and a series of primers 8-1R to12R (SEQ ID NOs: 22-33) as well as using pSA212 as a template. Each of the 12 types of the resulting DNA fragments were inserted at the KpnI-EcoRI site in the plasmid described above. The resulting vectors were used for the identification.